

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Kaare M. GAUTVIK et al.

Title:

Production of Human Parathyroid

Hormone From Microorganisms

Appl. No.:

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L. Spector

Art Unit:

1647

DECLARATION OF KAARE M. GAUTVIK, M.D.

PURSUANT TO 37 C.F.R. § 1.132

Mail Stop NON-FEE AMENDMENT Commissioner for Patents PO Box 1450 Alexandria, Virginia 22313-1450

Sir:

I, KAARE M. GAUTVIK, declare as follows:

- 1. I am a co-inventor of the above-captioned application.
- 2. I am a citizen of Norway residing at Bregnevn 3, 0875 Oslo, Norway. I am fluent in English. My curriculum vitae is attached hereto as exhibit A.
- 3. The standard hPTH(1-84) referred to on page 7, line 19, of the specification is synthetic hPTH obtained from chemical supply companies, including Peptide Institute Protein Research Foundation, Penninsula Laboratories, Sigma, and Bachem.
- I participated in the experiment detailed below to study the purity of the synthetic hPTH obtained from Peptide Institute Protein Research Foundation, Penninsula

Laboratories, Sigma, and Bachem as compared to recombinant hPTH produced according to the claimed invention. The experiment was performed on November 30, 1989 and the commercial hPTH preparations were purchased from 1986-1989.

Experimental Method

5. 0.2 μg of hPTH(1-84) obtained from Peptide Institute Protein Research Foundation, Penninsula Laboratories, Sigma, and Bachem, and from hPTH(1-84) produced according to the claimed invention, was analyzed on an SDS-PAGE gel. The protein bands were stained by silver nitrate. Silver nitrate is the most sensitive stain to disclose impurities. The hPTH(1-84) was loaded onto the gel as follows. Where available, lot numbers, catalog numbers, and date of arrival to the laboratory are provided.

Lane 1: BRL, low molecular weight standard

Lane 2: Peptide Institute Protein Research Foundation, lot no 350306, arrived July 1986

Lane 3: Penninsula Laboratories, arrived July 27th 1986

Lane 4: Sigma P-7036, arrived before October 1st, 1987

Lane 5: Bachem, lot no ZF567, arrived July 1989

Lane 6: Bachem, lot no 734B, arrived February 1989

Lane 7: E.coli PTH (recombinant) (produced according to claimed invention)

Lane 8: Yeast PTH (recombinant) (produced according to claimed invention)

Lane 9: Yeast (Q26) PTH (recombinant) (produced according to claimed invention)

Lane 10: BRL, low molecular weight standard

Results

- 6. A photograph of the SDS-PAGE gel obtained from the above-described experiment is provided as Exhibit B. Since the lanes of the gel were loaded with the same amount of hPTH(1-84), 0.2 μg, according to the manufacturer's description, the intensity/size of the bands should be the same. However, as shown in the SDS-PAGE gel picture attached as Exhibit B, the intensity/size of the bands is not the same for the hPTH(1-84) obtained from Peptide Institute Protein Research Foundation, Penninsula Laboratories, Sigma, and Bachem (lanes 2-6), as compared to the recombinant hPTH produced according to the claimed invention (lanes 7-9). Further, impurities (shadows appearing above or below the main band) are apparent in the lanes containing hPTH(1-84) obtained from Peptide Institute Protein Research Foundation, Penninsula Laboratories, Sigma, and Bachem, but are virtually absent in the lanes containing recombinant hPTH produced according to the claimed invention. A more detailed analysis of the results of the gel is provided below.
- 7. Lane 2: The molecular weight of the main band is higher than the expected molecular weight for hPTH(1-84). Impurities are seen on the photograph above and below the major band. The material in lane 2 contains less hPTH(1-84) than the manufacturer's stated quantity.
- 8. Lane 3: The molecular weight of the main band is higher than the expected molecular weight for hPTH(1-84). Impurities are seen on the photograph above and below the major band. The material in lane 3 contains less hPTH(1-84) than the manufacturer's stated quantity.
- 9. Lane 4: Low molecular weight impurities are present. The material in lane 4 contains less hPTH(1-84) than the manufacturer's stated quantity. The impurities lead to false weight estimations.
- 10. Lane 5: Low molecular weight impurities are present. The material in lane 5 contains less hPTH(1-84) than the manufacturer's stated quantity. The impurities lead to false weight estimations.

- 11. Lane 6: Low molecular weight impurities are present. The material in lane 6 contains less hPTH(1-84) than the manufacturer's stated quantity. The impurities lead to false weight estimations.
- 12. Lane 7: The hPTH(1-84) is more than 95% pure hPTH(1-84) of the correct molecular weight. There is virtually no material above or below the major protein band.
- 13. Lane 8: The hPTH(1-84) is more than 95% pure hPTH(1-84) of the correct molecular weight. There is virtually no material above or below the major protein band.
- 14. Lane 9: QPTH moves a little faster than hPTH(1-84) because of one amino acid residue substitution, and there is no sign of impurities
- 15. Although the synthetic hPTH (1-84) obtained from Peptide Institute Protein Research Foundation, Penninsula Laboratories, Sigma, and Bachem was impure, this synthetic hPTH (1-84) was used as a hPTH (1-84) standard in my lab. The impure hPTH (1-84) standard was used to confirm in early experiments the identity and characteristics of hPTH (1-84) produced according to the claimed invention. The impurities present in the synthetic hPTH (1-84) standard did not as such, affect the usefulness of the synthetic hPTH (1-84) as a qualitative "standard."
- 16. As a consequence of impurities and incorrect amounts provided by the commercial producers, the specific biological activity of even their best preparation was found to be about 30% below that of the recombinant hormone. (See, e.g., exhibit C, Reppe, S., et al., 1991, JBC, 22:14198-14201, Figure 5).

Atty. Dkt. No. 016777/0310 Appln. No. 08/340,664

17. I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true, and further that these statements are made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Dated: 07.27.64

Kaare M. Gautvik, N

CURRICULUM VITAE RELEVANT PUBLICATIONS KAARE M. GAUTVIK, M.D., PHD, CHIEF CONSULTANT

Personal and marital status:

Name:

Kaare M. Gautvik

Home address:

Bregnevn. 3, 0875 Oslo, Norway

Business address:

Institute of Medical Biochemistry, University of Oslo.

P.O.Box 1112 Blindern, 0317 Oslo, Norway

Telephones: Date and place of birth: 47-22851055 (work); 47-22235137 (home)

Social Security:

11th of December 1939 in Oslo.

No.: 111239.39311

Married to:

Vigdis Teig Gautvik, date of birth: 24th of March 1947

Children:

Lars Erlend Sakrisvold Gautvik, date of birth: 9th of January 1964

Silja Marie Sakrisvold Gautvik, date of birth: 31th of March 1973, Ole Martin Teig Gautvik, date of birth: 21th of

January 1982

Education and Clinical Specialities:

August 1958-June 64, Medical School at the University of Oslo.

- 1967-69, Courses in mathematics involving geometry, statistics and mathematical analysis.
- 3. May 1970, Disputation for the medical doctor degree at the University of Oslo.
- 4. 1985, Specialist in clinical chemistry, and physiology and nuclear medicine.
- 5. 4986, Specialist in occupational health medicine.

Employment:

- June 1964-June 1965, working at Tromsø University Hospital at medical and surgical departments.
- July 1965 until December 1965, working as a general practioner in Sjøvegan, Troms.
- One year military service as a major in The Norwegian Air Force. working mainly at the Norwegian Institute for Aviation and Space Medicine.
- From 1967, position as post-doctoral researcher at The Institute of Physiology, University of Oslo.
- From September 1969, promoted to Assistant Professor at the University of Oslo, Institute of Physiology.
- Leader and responsible for clinical and experimental endocrinological laboratory of Institute for Surgical Res., The National Hospital, Oslo, from 1973-89.
- 7. From 1976-1978, training as a specialist in clinical chemistry at the Norwegian Radium Hospital, Oslo.
- From August 1983 appointed to full professor at the Institute of Medical Biochemistry, Medical Faculty, University of Oslo.

(At the same time receiving offers of professor chairs at the Institute of Physiology, Medical Faculty and at the Institute of Physiology and Biochemistry, Faculty of Odontology).

From Jnuary 2002 employment as senior consultant at Department of Clinical Chemistry, Labaratory Division, Ulleval University Hospital and professor II at the University of Oslo.

Post-doctoral training abroad:

- For three months in 1967, I worked as a lecturer at the Department of Physiology, Medical School, Birmingham University, England.
- From August 1971 to July 1973 in receipt of Fogarty international post-doctoral fellowship at the Department of Pharmacology, Harvard School of Dental Medicine and Harvard Medical School.
- 1980, 4 months Visiting Professor at Institute of Genetics, BMC, Uppsala University, Sweden.
- 1995/96, 12 months Visiting Research Professor, The Scripps Research Institute,
 - Dept. Mol. Biology, La Jolla, San Diego, USA.
- 1997, 3 months Visiting Research Professor, The Scripps Research Institute, Dept. Mol. Biology. La Jolla, San Diego, USA.

Teaching responsibility:

- One year teaching in aviation medicine for medical personnel and pilots.
- I have given lectures and courses for medical students in following subjects:

Haematology, kidney physiology, endocrinology, circulation, respiration and gastrointestinal physiology.

From 1983 organized and given lectures and courses in molecular genetics at undergraduate and postgraduate level for students in medicine and sciences.

- Organized interfaculty advanced courses within molecular endocrinology.
- Lectures have been given in the following subjects at post-doctoral courses:

Diseases of the thyroid gland (1973); Regulation of circulation in the gastrointestinal system (1973); Local hormones (1975); Endocrinology (annually from 1978); Tumour markers (1979); Calcium metabolism (annually from 1980); Ligands for peptide hormonereceptors, and Nuclein acid biochemistry (1984); TRH-receptors in prolactin-producing cells (1985). Molecular biology in medical research (yearly from 1983). Biochemical analysis on bone material (1991).

Invited lectures: Several places in the U.S., in Sweden, in Finland, and in England, as well as different places in Norway, a total of 37 as of 1995.

- Chief organizer of post graduate scientific courses for the Medical Faculty at University of Oslo, 1986-1991.
- Organizer of international scientific meetings within the frame of the following societies:
 Acta Endocrinologica (European International Endocrine Society), The Scandinavian Physiology and Pharmacology Meetings, and the Norwegian Biochemical Society.
- 8. Introduced teaching in Molecular Biology for students at the Medical Faculty, Oslo.
- 9. Invited as Symposium Lecturer at international meetings in physiology and endocrinology and molecular biology as exemplified below:

Examples of specially invited symposium lectures:

- February, 1990: "Production of recombinant human parathyroid hormone in E.coli and Saccharomyces cerevisiae and its potential use as drug in osteoporosis" by Kaare M. Gautvik, Eli Lilly Co., Indianapolis, USA, in a Biotechnology meeting.
- June, 1990: Symposium lecturer and organizer: "Hormone receptors and cellular signal transduction. The XXII Nordic Congress in Clinical Chemistry, Trondheim, Norway.
- July, 1990: Symposium lecturer: "Transmembrane signal systems involved in the regulation of prolactin secretion by hypothalamic peptide hormones in cultured pituitary cells. 2nd European Congress of Endocrinology, Ljubljana, Yugoslavia.
- 4. July, 1990: Symposium lecture: "Successful cloning and production of human parathyroid hormone (hPTH) in yeast as a secretory product". 5th European Congress on Biotechnology, Copenhagen, Denmark. (Unable to attend, and the lecture was held by cand.scient. Sjur Reppe).
- August, 1990: Symposium lecture: "Processing and stability of human parathyroid hormone produced in E.coli and S.cerevisiae studied by in vitro mutagenesis". Workshop/Symposium on site-directed mutagenesis and protein engineering, Tromsø, Norway.
- 6. December, 1990: Invited by Professor Guo Hui-Yu, Guangzhou, China and Professor G.L. French, Hong Kong. Lecture entitled: "Expression of human-parathyroid hormone as a secretory protein in prokaryotic and eukaryotic microorganisms". The Second International Conference on Medical Microbiology and Biotechnology Towards 2000, Guangzhou, China. (Did not attend as a protest against the punishment of the students rebellion in Peking).
- January 1991: Invited to a Workshop by Dr. Stephen Green, Central Toxicology Laboratory, ICI, Alderly Park, Maccelesfield SK10 4TJ, UK. Lecture entitled: "Synergistic effects of hormones and fatty acid on peroxisornal 8oxydation, enzyme activities and mRNA levels".
- January 1991: Invited to a Protein Engineering Meeting by Professor Ian Campbell, Biochemistry Department, Oxford University, Oxford, UK. Lecture entitled: "Cloning and expression of human parathyroid hormone in microorganisms".
- Invited by Professors T.T. Chen, D.A. Powers, B. Cavari, Maryland Biotechnology Institute, Baltimore, MD, to held a symposium lecture at the 2nd International Marine Biotechnology Conference, October 13-16, 1991, Baltimore, Maryland. USA. (Could not attend).
- May 1991: Invited by Professor Jan Carlstedt-Duke, Karolinska Institutet, Huddinge, to held a lecture in the seminar series "Novum Lectures in Cellular and Molecular Biology".
- 11. January 1992: Invited by Professor Armen H. Tashjian, Department of Molecular and Cellular Toxicology, Harvard School of Public Health and Department of Biological Chemistry on Molecular Pharmacology, Harvard Medical School, Boston, USA. Lecture entitled: "Use of antisense RNA in delineation of the mechanism of action of G-coupled hormones".
- August 1993: Invited by Norwegian Society of Charted Engineers, The Blindern Conference. Lecture entitled: "Experience from industrializing basal research".
- 13. November 1993: Invited by Karolinska sjukhuset, Stockholm, to held a lecture at "Graduate course in molecular endocrinology a problem oriented approach". The lecture is entitled: "Region specific actions of parathyroid hormone in target tissues".
- 14. February 1994: Invited by GBF, Gesellschaft für Biotechnologische Forschung mbH, Braunschweig. Lecture entitled: "Expression of human parathyroid hormone in microorganisms and animal cells with special reference to signal sequence efficacy and intracellular modifications".
- 15. September 1994: Invited by Professor K. Dharmalingam, Department of Biotechnology, Madurai Kamaraj University, India, to held a lecture in the symposium "Gene expression systems", XVIth IUBMB, New Delhi. Lecture entitled: "Expression of human parathyroid hormone in microorganisms, insect cells, mammalian cells and as a milk protein in transgenic mice".
- 16. November 1994: Invited by Professor A. Taschjian Jr., Harvard School and Public Health, Boston, to held a lecture in a seminar. Lecture entitled: "Certain structural and functional characteristics of the human TRH receptor cDNA and mapping of the gene".
- 17. February 11-13, 1995: Cairns, Australia, Workshop on "Animal models in the prevention and treatment of osteopenia".
- 18. February 1995: Int. Meeting of Calcified tissue research, Melbourne, Australia.
- May 1996: Dublin University Program. "How to identify patients at risk for development of ostoporosis".
- September 1996: Lecture at Scripps Research Institute, San Diego. "Unique hypothalamic specific mRNAs expressed by molecular subtraction hybridization".
- September 1996: Invited seminar at the Astra Research Center, Montreal. "Cloning and expression of human polypeptide hormones with biomedical potential".
- November 1996: Invited lecturer, The Norwegian Rheumatological Society, Oslo, "PTH (parathyroidea hormone) -The biochemical foundation for treatment of osteoporosis".
- December 1996: Invited at Nordic Conference for Medical Treatment of Osteogenesis Imperfecta, Holmen Fjordhotel, Asker, Norway. "Characteristics of bone remodelling in patients with osteogenesis imperfecta".

- January 1997: Invited lecturer at The Salgrenska Hospital in Sweden. "Characterization and functional analysis of novel hypothalamus genes as identified by directional tag substraction".
- 25. 1998: Guest lecturer at Scripps Research Institute: "Hypothalamic calcium-calmodulin kinase-cloning and functional aspects".
- 26. February 1999: Only invited speaker from abroad at National Osteoporosis Congress in Rio de Janeiro, Brazil.
- 27. 2000: Lecture at NPS-Allelix company and Toronto University: "Parathyroid hormone regulated bone remodelling".
- 28. May 2000, Rio de Janeiro, Brazil. Member of the International Scientific Panel at the International Congress in Osteoporosis.

Honorary lectures and prizes:

- 1. In 1984 recipient of Professor Olav Torgersen's Prize and Memorial lecture. This prize and lecture was created by Professor Torgersen, the University of Oslo, who was one of the founders of the Society for Promotion of Cancer Research in Norway. Because he contributed with personal money, the prize and lecture had his name. The title of my lecture was: "The medullary thyroid carcinoma: a special type of familial and hormone producing cancer".
- 2. In 1984 I was given the international science prize called The Nordic Insulin Prize instituted by Professor Jacob E. Poulsen, who worked at the University of Copenhagen. This prize is given within endocrinology and the candidate is chosen from all the countries in Northern Europe. The money was donated by the Insulin Laboratory now the company Novo-Nordisk. At that time, only one Norwegian had previously received this prize. The prize was given for my studies regarding how hormones exerted their biological actions in target cells.
- The Gunnerus Prize was given in 1986 by the Royal Society of Norwegian Scientists. This is a prize which is given to a scientist selected by this society for scientific merits obtained and again it was within the field of hormone structure and action.
- In 1987 I received a prize within biotechnology created by the Research Park at the University of Oslo, which at that time was called the Innovation Centre, University of Oslo.
- 5. Novum Lectures in Cellular and Molecular Biology, which was associated with a scientific prize. Invited by Professor Jan-Åke Gustafsson at Novum, Huddinge, The Karolinska Institute, Sweden, in 1991. This was given based on my research with human parathyroid hormone in relation to its first cloning, expression and studies of actions.
- Lectures at Harvard School of Public Health in Cellular and Molecular Biology in 1995, regarding cloning of hormone genes and their characterizations. Invited by Professor A.H. Tashjian Jr. at the Department of Molecular and Cellular Toxicology, Harvard School of Public Health and Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, Boston, USA.
- 7. Given a 3 years economical "Group Research Support of 0.8 M NOK per year from 1997" after national and international project evaluations.
- 8. Scientific prize (Abstract award) 1997, at the Endocrine Society "Cloning and Organization of the human TRH-receptor Gene".
- 9. Norwegian Endocrine Society Prize (S.Reppe) for "Sox4 a PTH regulated transcription factor in bone".
- 10. Endocrine Society 1998: Poster Award: Cloning and regulation of the thyroliberin receptor gene.
- 11. Norwegian Endocrine Society prize 1998: Hormone regulated bone remodelling.
- 12. American Society for Bone and Mineral Research (ASBMR); 1999 Best Poster Award: The Transcription factor Sox-4 is expressed in developing cartilage and bone cells.

Consulting appointments:

- 1. Senior honorary consultant for NPS Biotechnology, Salt Lake City, Colorado, USA.
- 2. Consultant for Karolinska Institute, Stockholm, Sweden.

Referee activity:

I am or have been working as referee for the following international journals:

Endocrinology, J. Expl. Cell Res., Acta Physiol. Scand. (Kbh.), Eur. J. Endocrinol. (Acta Endocrinol. Scand. (Kbh.), Eur. J. Clin. Invest., Hormone Research, Acta Obstet. Gynecol. Scand., Journal of Endocrinological Investigation, Eur. J. Biochem., Experimental Cell Research, J. Biol. Chem.

Guidance for the academic doctor degree: Twenty three and 5 ongoing.

Supervision of postgraduate candidates: Presently three.

Supervision of students' main degrees: Nineteen.

Guest research workers from abroad: In my group we have had research visitors for periods of one to three years from Polen, Bulgaria, Sweden, Tyskland, Denmark, Iceland, India, Israel and USA.

Member of committees for the academic doctor degree in Norway and abroad: 15.

Member of advisory international/national committees for evaluation of professor positions:14.

Honorary Societies: Member of the Norwegian National Academy of Science and Letters

Professional memberships: Norwegian Society of Biochemistry, Norwegian Society of Physiology, Norwegian Society of

Endocrinology, Endocrine Society (USA), American Society for Bone and Mineral Research (USA)

Medical Faculty Responsibilities:

- 1. An elected member of the Medical Faculty 1987-1990.
- 2. A member of the Research Council at the Medical Faculty 1987-1990.
- 3. Chairman of Postgraduate Courses for Ph.D. and Dr.med. students at the Medical Faculty 1986-1991.
- 4. Member of the Institute Group Committee for the Preclinical Sciences from 1989 and present.
- 5. Member of the Medical Faculty's councel for evaluation of postgraduate applications from 1989-1993.
- 6. Committee member of the Medical Faculty's Scientific Instrument Board, 1996-.
- 7. Committee for Medical Research collaboration and interaction between University of Oslo and the National Hospital, 2000-.

National- and International Research Council Responsibilities:

1. Leader of Chemical Peptide Synthesis Core facility 1984-1989.

- Chairman for the Biotechnology Committee as a representative for Norwegian Research Council in an inter research council body, 1986-1989
- Member of The Norwegian Research Council for Science and the Humanities (NAVF) Committee for Physiology and Pharmacology, 1986-1989
- 4. Development and function as responsible leader of the nationwide core facility for peptide synthesis, 1988-1991.
- Member of the Premedical Institute Group Committee for Preclinical Sciences from 1989-2003.
- 6. Member of the International Scientific Board of Novo-Nordisk Research Committee, 1989-2001.
- 7. Member of the CIBA Foundation Scientific Advisory Panel from 1995 elected as representative from Norway, 1989-present.
- Chairman of the Research Council in the Norwegian Association for Osteoporosis, 1993-2003.
- 9. Leader of DNA Sequencing Core facility of the Institute of Basic Science, 1999-present.
- 10. Consultant and peer reviewer within Wallenberg Consortium North Technology Platforms DNA; SNP (single nucleotride polymorphism) Technologies and the Platform for Proteomics on behalf of the Board of the Wallenberg Consortium North, Stockholm, Sweden, 2001-2004.
- 11. Coordinator for Marie Curie Training Sites Fellowship No MCFH-200-00040: "Oslo Doctoral Training Site for Diagnosis and Therapy of Osteoporosis"—2001-2005.
- 12. Coordinator of EU 6.Program STREP contract no 502941, "Molecular mechanisms of bone homeostasis" (OSTEOGENE). Eight partners in 5 countries- 2003-2006

Awards and fellowships:

1967, 3 months, I worked as a lecturer at the Department of Physiology, Medical School, Birmingham University, England.

1971 to July 1973 in receipt of Fogarty international post-doctoral fellowship at the Department of Pharmacology, Harvard School of Dental Medicine and Harvard Medical School.

1980, 4 months Visiting Professor at Institute of Genetics, BMC, Uppsala University, Sweden.

1995/96, 12 months Visiting Research Professor, The Scripps Research Institute, Dept. Mol. Biology, La Jolla, San Diego, USA.

1997 Awarded for 3 years the Norwegian Research Council's Science Prize for outstanding research.

2001 Promoted by EU to become Oslo Doctoral Training Site for Diagnosis & Therapy Of Osteoporosis, received by a group consisting of scientists from University of Oslo, IMBA and the National Hospital.

2002 Member of CNS Molecular Biology group (leader Ivar Walaas) appointed as a "Research Theme Priority" at the Medical Faculty,2002- 2007

2004 OSTEOGENE (Molecular mechanisms of bone homeostasis) project given the highest priority and the only selected for presentation within Health Region East 2004.

Other professional activities:

- Founder of the Norwegian Association for Osteogenesis Imperfecta 1978 (Norsk Forening for Osteoporosis Imperfecta) together with Mrs.
 Myhre
- Founder of the Norwegian Association for Osteoporosis (Norsk Osteoporoseforening) 1993, together with Norwegian Women Public Health Association (NKS).

Patents:

I.Two U.S. patents, U.S. Patent No. 5.010 010 and No. 5.420.242 are held with international extensions in Europe, Japan, Canada, and Australia. In addition, three Divisional Applications are submitted to the U.S. Patent Office and elsewhere. These patents and patent applications in the different countries are covering specific methods related to the production, purification and characterization of PTH in microorganisms for the use in treatment of osteoporosis.

II.Inventor in patent application from Scripps Research Institute on: Novel hypothalamic mRNAs, the corresponding peptides and their functions.

Publications: More than 200 original articles published in internationally well reputated and refereed journals. Relevant articles are cited in relation to description of the research activities:

A BRIEF DESCRIPTION OF THE MAIN RESEARCH PROJECTS AND RELEVANT REFERENCES

A.STUDIES OF HYPOTHALAMIC SPECIFIC mRNAs OBTAINED BY A NOVEL SENSITIVE SUBTRACTION HYBRIDIZATION PROCEDURE

The results so far from our refinement and usage of a powerful and highly sensitive novel subtractive nucleic acid hybridization method have been successful. The generated hypothalamic subtraction library appears to give a specific and comprehensive representation of mRNAs that are not present in other brain areas as hippocampus and cerebellum. We have so far described several novel peptides: hypocretin (the cause of Narcolepsy) and several very interesting peptides, e.g. novel CaM kinase (see list of references). Another CNS peptide is somatostatin-like, called cortistatin, structure similarity with somatostatin; P25 and Vat 1, two uniquely expressed peptides in distinct regions of the brain.

1. Gautvik, K.M., de Lecea, Luis, Gautvik, V.T., Danielson, P.E., Tranque, P., Dopazo, A., Bloom, F.E. and Sutcliffe, J.G. Overview of the most prevalent hypothalamus-specific mRNAs identified by directional tag PCR subtraction.

Proc. Natl. Acad. Sci. USA (PNAS) 93: 8733-8738, 1996.

2.de Lecea, L., Criado, J.R., Prospero-Carcia, O., <u>Gautvik, K.M.</u>, Schweitzer, P., Danielson, P.E., Dunlop, C.L.M.Siggin G.R., Henriksen, S.J. and Sutcliffe, J.G. A cortical neuropeptide with neuronal depressant and sleep-modulating properties. Nature 381: 242-245, 1996.

3.de Lecea, L., Kilduff, T.S., Peyron, C., Gao, X-B., Foye, P.E., Danielson, P.E., Fukuhara, C., Battenberg, E.L.F., Gautvik, V.T., Bartlett II, F.S., Frankel, W.N., Van den Pol, A.N., Bloom, F.E., <u>Gautvik, K.M.</u> and Sutcliffe, J.G. The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. Proc. Natl. Acad. Sci. USA (PNAS) 95: 322-327, 1998.

B. ISOLATION AND CHARACTERIZATION OF mRNAs SPECIFIC FOR THE BONE CELL PHENOTYPE OBTAINED BY SUBTRACTION HYBRIDIZATION

By using the same novel subtractive hybridization procedure as employed and described above, we have generated a subtracted cDNA library using the osteosarcoma phenotype cDNA library as made from three different human osteosarcoma cells from which is subtracted the cDNA library obtained from normal human osteoblasts. The subtraction is performed by using cDNA from osteosarcoma cells minus RNA transcribed from the corresponding cDNA library of the normal osteoblast. These are experiments in progress and we are about to describe individual cones obtained from a subtracted library of about 400.000 independent colonies. The aim of this study is to identify those mRNAs which are overexpressed or lacking in the osteosarcoma phenotype and compile these results in order to have a greater understanding regarding how a normal cell is transformed into this tumor type (Olstad et al., 2003)

C.ISOLATION AND CHARACTERIZATION OF mRNAs SPECIFIC FOR PARATHYROID HORMONE GENE ACTIVATION IN BONE CELLS ("Bone anabolic genes")

Parathyroid hormone is the most important physiological regulator of bone formation. This hormone therefore is assumed to represent an important drug in the prevention and especially treatment of postmenopausal osteoporosis. However, as a succession of our previous work regarding the studies of this hormone, we have continued to search for a complete overview of all gene products that parathyroid hormone is stimulating in bone cells in order to isolate the mRNAs and corresponding proteins which may be of sentral importance for the development of osteoporosis - or which may be called "the genes for osteoporosis". Again by using the same molecular substraction method as described in Chapter II, C, we use this time parathyroid stimulated normal bone cells cDNA library minus RNA transcribed from generated libraries of normal bone cells. This work is almost completed in a highly successful manner. We have isolated more than 40 genes which are involved in PTH anabolic action in bone, and among those we are searching for the gene(s) causing postmenopausal osteoporosis.

D. As a complementation and to the activities described above, we have embarked on defining the bone phenotype in female and male osteoporosis within the context of the EU project OSTEOGENE (see above). About 100 patients and controls will have bone biopsies which will be prepared and analysed for their global gene expression and differences at the micro- and ultrastructural level. I am the coordinator of this activity including 5 countries and where Oslo university and three hospitals (Ulleval university hospital, the National Hospital and Lovisenberg hospital are working closely together. This is a direct consequence and follow up of previous research representing patient related basic and translational science aiming to solve the mechanisms of osteoporosis, the most common disease in women of 50 yrs of age.

THE MAIN RESEARCH ACTIVITIES DURING THE LAST 8 YEARS AND FUTURE SCIENTIFIC ENGAGEMENT:

PARATHYROID HORMONE (PTH) AND PARATHYROID HORMONE RELATED PROTEIN (PTHrP)

The aim for this work was to produce:

- i) Recombinant parathyroid hormone for structure activity studies in relation to bone cell activation.
- ii) Study intracellular processing and trafficking of these hormones and to compare signal sequence efficacy in different host expression systems.

We were the first in the world to clone and produce full-length <u>human recombinant parathyroid hormone</u> in mg quantities. For this work we developed gene constructs, vector modifications, fermentation technological improvements as well as complete methods for down-stream technology. The final product is PTH identical and more than 99% pure and has shown full chemical, biochemical and biological identity with the intact hormone. These results are written in the following articles that are printed.

We have also been as indicated by the list of references below, the first in the world to express secreted human parathyroid hormone in mammalian cells as well as a secretory milk product in transgenic mice. In addition, we have been the first to develop full-length PTH polypeptides with agonist and antagonist functions.

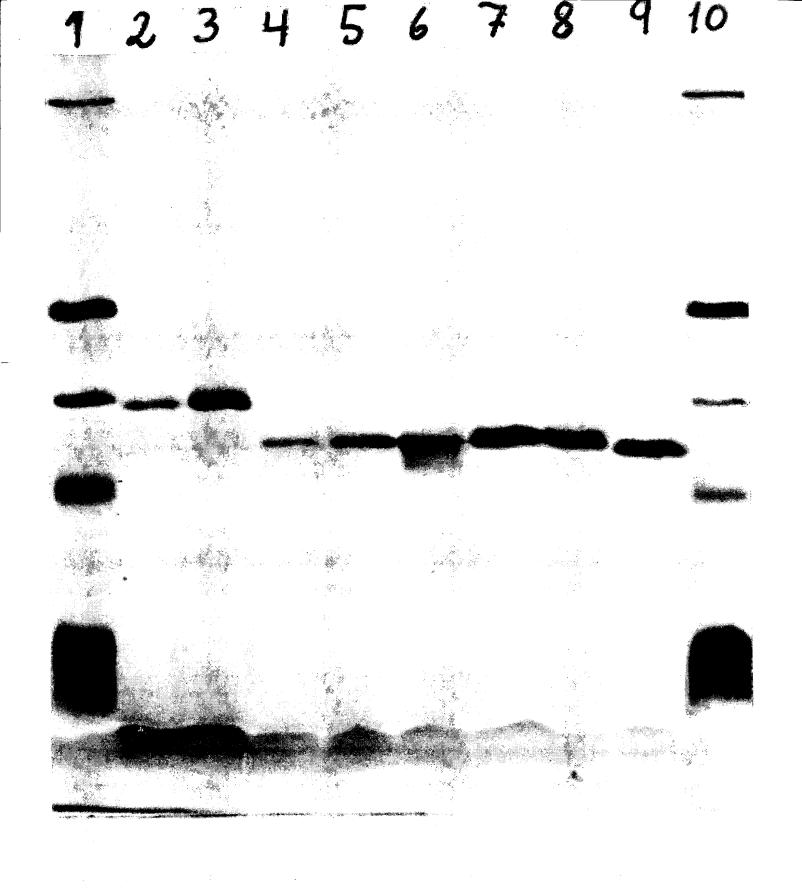
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Characterization of a K26Q Site-directed Mutant of Human Parathyroid Hormone Expressed in Yeast*

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From the ‡Department of Biochemistry, University of Oslo, P. O. Box 1041 Blindern, 0316 Oslo 3, Norway, the §Institute of Medical Biochemistry and Biotechnology Centre, University of Oslo, P.O. Box 1112 Blindern, 0317 Oslo 3, Norway, the ¶Institute for Surgical Research, The National Hospital, University of Oslo, Pilestredet 32, 0027 Oslo 1, Norway, the **Center for Industrial Research, P. O. Box 124 Blindern, 0314 Oslo 3, Norway, and the ‡‡Laboratory of Toxicology, Harvard School of Public Health and the Department of Biological Chemistry and Molecular Pharmacology, Harvard Medical School, Boston, Massachusetts 02115

Human parathyroid hormone (hPTH) is susceptible to proteolytical cleavage both in humans and when expressed as a secretory product in Escherichia coli (Høgseth, A., Blingsmo, O. R., Sæther, O., Gautvik, V. T., Holmgren, E., Hartmanis, M., Josephson, S., Gabrielsen, O. S., Gordeladze, J. O., Alestrøm, P., and Gautvik, K. M. (1990) J. Biol. Chem. 265, 7338-7344) and Saccharomyces cerevisiae (Gabrielsen, O. S., Reppe, S., Sæther, O., Blingsmo, O. R., Sletten, K., Gordeladze, J. O., Høgset, A., Gautvik, V. T., Alestrøm, P., Øyen, T. B., and Gautvik, K. M. (1990) Gene (Amst.) 90, 255-262). In the latter system, one major site of cleavage was identified (Arg25-Lys26 Lys27). To produce hPTH resistant to this proteolytic processing, a point mutation changing Lys26 to Gln was introduced. and the modified gene expressed in S. cerevisiae as a fusion protein with the α -factor leader sequence. The resulting major form of hPTH secreted to the growth medium was of full length showing that the mutation had eliminated internal processing. Consequently, the yield of the mutant hormone was significantly higher than obtained with the natural peptide. Using improved purification procedures, a significantly higher purity was also obtained. The secreted mutant hPTH-(1-84,Q26) had the correct size, full immunological reactivity with two different hPTH antisera, correct amino acid composition and N-terminal sequence, and correct mass as determined by mass spectrometry. Furthermore, the introduced mutation did not reduce the biological activity of the hormone as judged from its action in three biological assay systems: 1) a hormonesensitive osteoblast adenylate cyclase assay; 2) an in vivo calcium mobilizing assay in rats; and 3) an in vitro bone resorption assay.

Gene fusions with the mating pheromone α -factor leader sequence have been widely used to express, correctly process, and efficiently secrete human proteins of medical interest in Saccharomyces cerevisiae (1-4). Contrary to the situation in higher eukaryotes where the propeptide processing signal seems to involve proximal residues in addition to the doublet of basic amino acids (5, 6), there is no evidence that yeast vscF endopeptidase (KEX2 gene product) requires more than a pair of basic amino acids to process a propeptide, although a Lys-Lys pair is a poor substrate for the enzyme (6, 7). Since the probability of finding two consecutive basic amino acids (Lys-Arg, Arg-Lys, or Arg-Arg) in a random protein sequence is rather high, many heterologous proteins risk to be aberrantly processed during secretion in yeast. Thus, in several cases where such sites have been present in the mature peptides, the secretion products were reported to be either partially (8) or completely (9) cleaved, heterogeneous (10), or to contain extra unidentified peptides (11), suggesting aberrant processing.

The mature human parathyroid hormone (hPTH)¹ illustrates this problem because it contains two potential processing sites (Arg-Lys) in the 84-amino acid-long molecule. Hence, when the hPTH gene, fused to the α -factor leader, was expressed in S. cerevisiae, a significant fraction of the synthesized protein was cleaved at the first of these sites. The cleavage site was localized after the two basic amino acids (Arg²⁵-Lys²⁶) Lys²⁷), suggesting that the yscF protease is involved (12).

In the present report we describe the production in yeast of a mutant hPTH where the internal putative yscF cleavage site has been removed by in vitro mutagenesis, substituting lysine at position 26 (Lys²6) by glutamine (Gln²6). As a consequence, the major degradation product hPTH-(27-84) (12) was no longer found in the growth medium while the yield of full-length hormone increased. The secreted hPTH-(1-84,Q26) had correct size, mass, sequence, and immunoreactivity. Furthermore, the introduced mutation did not reduce the biological activity of the hormone as judged from its action in several biological test systems.

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¹ The abbreviations used are: hPTH, Human parathyroid hormone; hPTH-(1-84), full-length (84 amino acids long) human parathyroid hormone with no mutations; hPTH-(1-84,Q26), full-length human parathyroid hormone where amino acid Lys²⁶ is mutated to Gln²⁶; prefix y, produced in yeast (S. cerevisiae); prefix s, synthetic (commercial preparation).

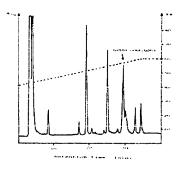
FIGURE 1.

*** 18.4 E

SDS PAGE of concentrated years growth medium containing mutated and non-mutated hPTII. Aliquots of concentrated growth medium from years strain 831991 transformed medium from yeals strain 531991 transformed with the expression plannids priUNPTH-Q16 (lane 3) and poiUNPTH-2 (12) (lane 2) were analyzed by 15% PAGE in the presence (11% SDS), and visualised by silver statisting as described in Materials and Methods. Lane 6: moticular size markers.

Lane 1: a commercial bPTH standard (Barkem).

FIGURE 2A.



HPLC mutification of senst sterived hPTHi1.84.12(a). Years medium concentrated on S Sepharose FF was further purified as described in Materiath and Methods. The elution profile from the first HPLC step, vigate CI8 culum, its abovan. The peak containing shFTHI1.84.(2)(a) is marked, Several peaks probably represent non-peptide low molecular wright aubstances from the fermentation broth since they do not give true to bands when analysed by SDS-PACE.

FIGURE 2B.

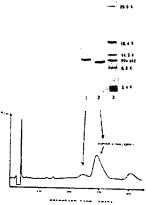


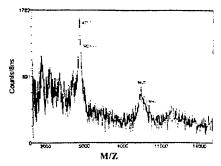
FIGURE 2B.

The elution profile from the second HPLC step, a Waters Nova-Pak C12 column, issueratic elution, with an analysis of the final purity of JMP William States and a silver staining. Lane 2 shows JMPTIII (184, (2)a) from the main HPLC peak. The molecular weight marker in lane 3 is, the same as in Figure 1, but includes a reference bPTII produced in E.c.aci. Lane 1 shows a slower migrating hormone (0.6 mg) which is Ogycosylated (umpublished results, and that elutes in the minor front peak from the second HPLC.

See Materials and Methods for experimental details Table 2 Tital amino acid composition of yhPfH(1-84,Q26).

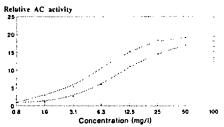
Λa	yhPTH(1-#4,Q26) Found Theory		hPTH(1-84) Found Theory		shPTH(1-84) ¹ Found Theory		
Ass	10.4	10	10.3	10	10.4	10	••••••
Gia	11.R	12	19.7	11	10.X	11	
Ser	7.4	7	7.3	7	7.4	7	
Gb	4.6		1.4	4	4.5	4	
His	3.7	4	3.8	4	3.7	4	
Thr	1.2	1	1.2	1	1.1	1	
Ala	6.9	7	4.7	7	6.8	7	
Arg	5.3	5	5.4	.5	5.4	5	
Pro	3.3	3	3.2	3	3.0	3	
Val	7.5	8	7.8		7.7	ä	
Met	1.8	2	1.8	2	1.7	2	
lte	0.9	1	0.9	1	0.8	1	
l.ru	9.5	10	9.6	10	9.7	0.1	
Phe	1.2	1	1.1	1	1.1	1	
Lyx	7.6	8	8.7	9	8.8	•	
Τm	nd.	1	æd.	1	nd.	1	
Sum	83.1	84	X2.9	84	82.9	84	***************************************
		tandard hPII		erimental	details.		

FIGURE 3.



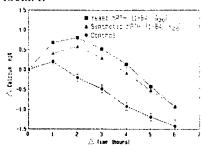
Plasma desorption mass spectrometry analysis of purified recumbinant shPTH(1-34.4126). The spectrum thems the on abundance as a function of mass over charge.

FIGURE 4.



Adenylate exclase-stimulating acitivity of vhPTH(1-84.016) Partified recombinant PFTH1-84(Q26) L.-.) was assayed for biological activity in a hormone-sensitive outcobast adenyiate cyclase (AC) assay as described in Materials and Methods, Relative AC artivity means relative to control-activity set equal to 1. Each point gives the mean raise of triplicate samples, hPTH(1-84) from Bachem (LL) was used as reference.

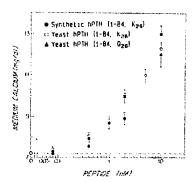
FIGURE 5.



la stee induction of hipercultemia by hPTI(13-84,02a)

Parahterodectionized male Weige cats were texted for induction of hypercultemia by the production of hypercultemia by the production of t

FIGURE 6.



B. ritro home renoration-atimulating activity of shPTH11-84(O)6)
Dose response relationships for shPTH11-841, yhPTH11-841, and yhPTH11-84,Q20) as hone recorption-atimulating agents. Each point gives the mean value and the brackets give the SF, for groups of S to 28 hones. Results were obtained from up to 5 independent experiments and were posted where statistically raild. The medium calcium concentration from control hones was 7.3 ± posites where Mainstreamy value, the inscrime continuous continuation in the Book of the B

MATERIALS AND METHODS AND RESULTS²

DISCUSSION

The hPTH appears to be especially sensitive to proteolytic degradation. Large amounts of hPTH fragments are found in the parathyroid gland (13), and only about 10% of circulating hPTH immunoreactive peptides are full-length hormone (14, 15). Similarly, degradation of hPTH has been a major problem when the hormone is expressed in heterologous organisms (16, 17). Improved yields of intact hormone have been obtained by secretion of the hormone into the growth medium, either in Escherichia coli (18) or in S. cerevisiae (12) or by expressing hPTH as an intracellular fusion protein in E. coli (19, 20). Even in these systems, proteolytically cleaved forms may be produced reducing the yield of intact hormone.

The present results show that hPTH can be expressed in a correctly processed and secreted in a largely intact form in S. cerevisiae after the introduction of a single, structurally conservative mutation in the 26th amino acid (Lys²⁶ \rightarrow Gln). The hormone produced resists the frequent cleavage found in the non-mutated hormone (12).

The increase in final yield from shake flask cultures of pure full-length hormone agonist is 5-10-fold compared with that obtained with the nonmutated hormone expressed in the same system (12) using identical culture conditions (results not shown). An improved purification procedure gave homogeneous hPTH-(1-84,Q26) after two high performance liquid chromatography steps.

The mutation was introduced in a region of hPTH previously found to be essential for biological activity (13) and within a triplet of basic amino acids conserved between the rat (21), porcine (22), bovine (23), and human hormone (24, 25). However, no difference between the activity of recombinant yhPTH-(1-84,Q26) and standard shPTH-(1-84) could be detected in three different biological assays: an adenylate cyclase assay, measurement of the ability to induce hypercalcemia in parathyroidectomized rats, and the mouse calvarial assay for bone resorption-stimulating activity. It is possible that the dibasic site in hPTH in fact functions as an easily attacked proteolytic site that destabilises the hormone and thus allows more rapid fluctuations in the hormone levels. A more degradation-resistant hPTH agonist could therefore be of potential importance when used as a medical drug due to different pharmacokinetics.

hPTH is one of the primary calcium-regulating hormones in the body and acts principally on kidney and bone cells, stimulating renal calcium reabsorption, phosphate excretion, and bone remodelling, respectively (26-28), resulting in anabolic as well as catabolic effects. Its overall physiological action is probably to generate a positive calcium balance and enhance bone formation. A fragment of hPTH together with 1,25-(OH)₂ vitamin D₃ have also been reported to induce bone formation in humans (29, 30), and one of the major areas of potential use of a recombinant hPTH is therefore in the treatment of postmenopausal osteoporosis. To evaluate such applications, sufficient supplies of recombinant hPTH or its agonists are essential.

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² Portions of this paper (including "Materials and Methods," "Results," Figs. 1-6, and Tables 1 and 2) are presented in miniprint at the end of this paper. Miniprint is easily read with the aid of a standard magnifying glass. Full size photocopies are included in the microfilm edition of the Journal that is available from Waverly Press.

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Supplemental material

to

Characterization of a K26Q site directed mutant of human parathyroid hormone expressed in yeast

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MATERIALS AND METHODS

Plasmids, strains and culture conditions.

Plasmids, strains and culture confidens.

The Saccharempers crientizes strain used for hPTH expression was BJ1991 (o. up1, ural-52, feul, pr31-1122, psp4-3). Yeast cells were transformed by the lithium method (31), and transformants grown at 30°C in VNBGC medium (0.67% yeast airrogen base, 2% glucose, 1% cassumbon section (Difro)). The putXFTH-2 plasmid used as a reference for expression of authentic hPTH(1.64) in yeast, and the subctone MISFTH-3, have been described previously (12; US Patent application on 251664).

In rare, mutagenesia on los o-lactos By In Imison gents.

The inappropriate processing of by PH secreted from yeast (cleaved at the site: Arg23.Lys264Lys27 (120)), might be eliminated by a change from Lys26 to Gla26 to destroy the potential yef processing site. To introduce this change, the subclose M13PTH-3 was modified by in ritro mutagenesis using the "Muta-gens" in vitro mutagenesis hit" (Bio-Rad) based on the method of Kanadel et al., (32) and the mutagenting off-genuleotide: "SCGTTGGTTCAGAACCTGC3." Positive plaques (containing M13PTH-Q26) were verified by DNA sequencing (33). The entire expression casettle between a Samili and a filled-in Scall site was isolated from M13PTH-Q26 and inserted between the BaseH1 and Prail alte of the yeast shuttle vector YEp24 (34). This mion plasmid was designated puUXPTH-Q26 and med to transform the yeast strain BJ1991.

Direction of APTH in culture media.

Raddolumanionamy of APTH in custome media was performed as described (12,35). For electrosphortic manyini, yeast culture media were concentrated on S Sephaross FF as previously described (12), and separated on a 15% polyacrylamide get in the presence of SIS (56), and either stalled with siture (37) or analyzed further by protein botting using immaphion PVDF Transfer Membrases (Millipore) and the buffers of Towbin et al (38). Reference hFTH(1-44) was purchased from Peninsula Laboratories (USA) and Sigma (USA). Protein blots were visualised as described

Parification of hFTH from yeast culture medium. The clarified culture medium was concentrated on a S Sephanose Fast Flow column previously couldinated with 0.3M gyicins, pH 3.0. The medium was adjusted to pH 3.0 with 1 M HCl before loading. The column was then washed with 0.1M NayHPO., pH 8.5. HFTH content with the Agg peak. This stap also resulted in a partial purification of the hormone. Final purification was obtained by a two-step reverse phase HPLC procedure. In the first step a lydac protein/peptide C18 column (2504.6 mm, purchased from The separation group*, Hesperia, CA,USA) was employed with a gradient from 40-60% channt B (claunt 8: 0.2% trifluoroacetic acid in 70% acedomitrile and 30% milliQ water, and claunts 8: 0.2% trifluoroacetic acid in 70% acedomitrile and 30% milliQ water, and clausts 8: 0.2% trifluoroacetic acid in 70% acedomitrile and 30% milliQ water, and essent a million in the second step, a Water Nova-Fak C18 column (10.0h.28 cm) was used with isocratic claution using 47% clusin 8. Both duates were monitored by UV at 220 nm.

N-terminal senosnoe. Amino acid composition and mass spectrometry.

Proteins to be sequenced were purified either by HFLC as described above or by SDS polyacrytamide gelelectrophoresis followed by blotting onto immobilion PVDF membranes (39). Automated Edman degradation was performed on a 471A Protein Sequencer with an on-line 120A pleayliblohydantoin amino acid analyzer from Applied Blosystems (Foster City, CA, USA). All reagents were obtained from Applied Blosystems. The amino acid composition was analyzed on a Biotronic SP4100 amino acid analyzer after hydrolysis of 30 µg of the hPTH peptide in 6M HCi

(special purity), 18.05% thiogycol acid, for 24 hr at 115 °C.

The molecular many of purified recombinant hPTH(1-84,Q26) was determined by 152Cf plasma described (18), and was kindly performed by Dr. Maris Harimanis, KabiGen AB, Stockholm.

Advantate crease assay for APTH. The adeaplate creases simulating activity of the recombinant hPTH was assayed as previously described using VMR 106 osteosarcoma cell membranes (12,46,41). Synthetic hPTH(1-84) from Signa was used as reference standard.

Assay of AFTH-induced hypercalcemia in rats Male Wistar rats (185-200 g) were parathyroidectomized using electrocautery 18 hours prior to the start of the experiment. The animals were fested oversight, and ansesthetized the next day using Hyporom Dornétum (0.2 mbras). The carected artery was cammutated using polyethylene-50 tuoting. The cannels was connected to a syringe containing Ringers Acctain, 4% bovins albumin, 32 Units heparin/ral. Five minutes after injection of 280 µl of the heparinized Ringera, a baseline blood sample was drawn (300 µl). The animals were trachestomized to prevent respiratory failure due to damage to the recurrent laryngest nerve running through the thyroid gland. PTH was then injected subcutaneously in the neck in a volume of 280 µl. Both PTH(1-84) and hPTH(1-84Q26) were dissolved in 100 µl 0.01N seetle acid. After dissolving, the solutions were brought up in 900 µl of Ringers Acetate containing 1% borine albumin. Blood samples were frawn from the caretid artery at 1, 2, 3, 4, 5, and 6 hours after the injection of PTH. The rais were re-begarinated 5 minutes is 1, 4, 5, 4, 5, and a nours after the injection of First. Lee "sax worn" re-experienced a minimal before drawing each blood sample using 100 µ in of the beparinized Ringers solution. The concentrations of FTH used were determined by amino acid ensitysis. bFTM(1-84,Q26) was used at 2µglanima, and the Backen reference FTH was used at 2.75 µg/animal (a higher nemerical value used to compensate for a discrepancy between what was labelled on the Backen vial (1 mg FTH), and what we determined by amino acid analysis (8-75 mg FTH)). The blood samples were centrifuged for 18 minutes, and the plasma was analysed for calcium using a Cobus Autoanalyzer (Cobas Bio, Rocke Disproaction).

Rose resormion-stimulating activity

Bose resormion-stimulating activity was measured using secondal mouse culvaria as previously
described (4.24). The medium was Delbecco's Medified Engle's medium (DMEM) supplemented
with 15% best-inactivated (54-69% C for 30 min) borns serum. Incubation of calvaria from 4 to 5day-old mice was performed at 179% in a roller drum apparatus under an atmosphere of 39% O₂.
5% CO₂, and 43% N₂. Calvariae were preincubated for 24 bours before experimental treatment 5% CO₂, nod 45% N₂. Calvariae were preinculated for 24 hours before experimental treatment was begun. The medium was then changed, and fresh control medium or medium containing standard synthetic FTH [shPTH(1-44)], prast derived hormone [yhPTH(1-84)], or yeast derived motiant hormone [yhPTH(1-84,Q26)] was added. Bone recorption was determined by measuring the accumulation of 40Ca2* in the culture medium during the 71-hour recalment period. The concentration of total calcium in the bone culture medium was measured in 400 µl samples by a calcium-selective electrode using a Nov 7-7 automatic calcium analyser (NOVA Blomedical, Waltham, MA, USA). Results in each experiment were subjected to an analysis of variance, and the SE values were calculated from the readule error term of that malysis. Experiments were posted when the variances were nonbeterogeneous. The shPTH(1-84) standard was from Bachem (Lot no. 1764.89).

RESULTS

RESULTS

Entraine and secretion of APTHIL-M.(Dis) in vess.

To prevent the internal processing of APTH occuring during secretion in yeast, a point mutation to prevent the internal processing of APTH occuring during lysin-26 to gittamine in the populous mutation was introduced in coden 26 of the APTH gave changing lysin-26 to gittamine in the populous mutation gave was expressed in Saccharenapers secretized to the growth sensition were studyes and the processing approaches and the processing approaches the standard APTH, and microscopencing showed that it corresponded to the hormous fragment APTH/2-64) (12). After mutation (lane 3), this band was absent, showing that the cleavage between amon code 26 and 27 was clinicated as a result of the incrossect mutation. The major product was a polypoptide that migrated close to the full length APTH standard, slightly faster in an anomale gil system (Fig.) and slightly slower in a cationic get system (act shown), in accord with the single charge difference between the mutated and the wild type proteins, in addition to the major product also migrated close to the APTH standard when analyzed by two dimensional get electropheresis (first dimension) arctic action/was given and shown) and, by protein blotting, it was found to react with two APTH specific analbodies against the 1-18 minos acid and 44-58 mino acid respectively (not shown), apporting that we are design with a full length hormone.

Parification
To parify yPTH(1-84,Q26), growth medium was first concentrated using a S Sephanose FF column (yielding 78% pure yhPTH(1-84,Q26)) (Fig.1). The bPTH peak from the first HPLC step contained predominantly yhPTH(1-84,Q26) but also some glycosysted bormone with reduced migration is SDS PAGE (Fig.1a). After a second sourceits HPLC chromotography, out y a single band was seen after SDS PAGE (Fig.28). When the yield from stable-flash cultures of pure yhPTH(1-84,Q26) was compared with that of yhPTH(1-84,45. I-6) float higher yields were obtained, consistent with our previous eximate of the fraction of full length bormons (up to 20%) produced with an expression plasmid encoding non-mistated hPTH(1-84) (12).

Biochemical characterization of whTH(1-84-026) The asters of the phFTH(1-84-026) The asters of the phFTH(1-84-026) The asters of the phFTH(1-84-026) product was verified by N-terminal amino seld sequencing showing correct amino-terminal sequence (Table I). In particular, the expected change from Lys to Gin in position 25 was confirmed. The amino acid composition of the purified phFTH(1-84-026) is shown in Table 2. Except for one Try that is not revested under the conditions used, the expected composition and total number of amino seld residuous were found. Therefore, no C-terminal degradation had occurred. To substantiate that the purified yhFTH(1-84-026) represented the instant hormone, mass spectrometry was performedry giving a plasma description mass spectrum se shown in Fig.3. A molecular mass of 94/2029 dations could be calculated from the single-charged and domble-charged molecular into present in the aperture. This value corresponds weed with the beautiful of the production of the

<u>Riological characterization</u>
The biological activity of the ybPTH(1-84,Q26) was tested by three different amays. The purified ybPTH(1-84,Q26) was first analysed for its ability to stimulate the adeaylate cyclase activity in rat UMR 106 cateonarcoma cetts (40,41) (Fig.4). The stimulation curve coincides with that of purified non-mutulate ybPTH(1-84). In this acute assay, no significant differences in biological activity was detected between the native hormone and the degradation-resistant mutated hormone.

Secondly, the purified yaPTH(1-84,Q26) and a standard shPTH(1-84) were assayed for the shifty to laduce hypercalcenia in parathyroidectomized rais (Fig.5). The yhPTH(1-84,Q26) had an in vive biological activity stimulatory effect comparable to or slightly above that of the synthetic hPTH central.

Finally, the yhPTH(1-M4,Q26) was tested for direct activity on hone by analying its bone recognition-stimulating activity using secontal mouse calvaria, yhPTH(1-M4,Q26) stimulated bone recognition in vitro- with a potency similar to that of abPTH(1-M4) and yhPTH(1-M4) IFI,6.5. A time-charge experiment similarity showed that yhPTH(1-M4,Q26) had the same bone recognition-stimulating activity as the non-mutated bormone (not shown). Thus, we have found no indication of reduced biological activity resiting from the introduced mutation.